

# **Bioinformatics Practical**

## **BS220 Medical Genetics**

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### **Objectives**

Understand DNA sequence alignment and its applications for medical problems. Learn how to use online tools to map a DNA sequence to the human genome and to multiple bacterial genomes using BLAST. Familiarise yourself with the database Online Mendelian Inheritance in Man (OMIM).

### **Story plot**

A patient is in a hospital in a critical condition. Medical doctors have extracted some pieces of DNA or RNA from patient's blood and have to decide, as a matter of life or death, what's going on with the patient. The patient is being treated for a genetic disease, cystic fibrosis, but the current symptoms cannot be simply explained by this diagnosis. In this situation, in addition to classic tests, a new test has been performed: all nucleic acids extracted from patient's blood plasma have been sequenced. You are provided with two sequences resulting from this experiment: sequence A and sequence B. You have two hours to analyse these and decide what these sequences mean for the patient's medical condition and how to save life.<sup>i</sup>

## Introduction

The **Story Plot** and the **Plan of the Practical** above contain several important terms, and before we proceed let's make sure you understand their meaning:

*Genetic disease* is a genetic problem caused by one or more abnormalities formed in the genome. Some of them are caused by *Mendelian inheritance*.

*Cell-free DNA* consists of degraded DNA fragments released to the blood plasma (the liquid part of blood that does not include blood cells). cfDNA pieces can come from apoptotic (dead) cells from all parts of the body. Human blood plasma normally should not contain any foreign DNA, only the DNA from the dead cells of this organism. E.g. if bacterial or viral DNA or RNA are present in blood, it may indicate infection and even sepsis.

*Sequencing* is the experimental procedure of determining the nucleotide sequence in DNA.

*Alignment (mapping)* is the process where you *align (map)* a given DNA sequence to some other DNA sequence. For example, you could compare a single short sequence to the long sequence of the human genome (~3 billion nucleotides), and ask a question, whether the human genome contains regions that have the same (or similar) sequences as our sequence of interest. If such region(s) exist in the human genome, then you can ask where these regions are located, and which of these regions better matches to our sequence of interest.

Task 1. Map sequence A to the human genome using BLAST

Here is “sequence A”:

AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGAAGAATTTTCATTCTGTTCTCAGTTTTCTGGA  
TTATGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTCCTATGATGAATATAGATACAGAAGCGTC  
AAGCATGCCAACTAGAAGAGGTAAGAACTATGTGAAAACTTTTGATTATGCATATGAAC

1.1. Let's go to the BLAST web site: <https://blast.ncbi.nlm.nih.gov>

The screenshot shows the BLAST website interface. At the top, there's a section titled "Basic Local Alignment Search Tool" with a brief description of BLAST and a "Learn more" link. To the right, there's a "NEWS" sidebar with a message about a new version of IgBLAST (1.17) and a date stamp. Below this, the "Web BLAST" section features three main options: "Nucleotide BLAST" (nucleotide to nucleotide), "blastx" (translated nucleotide to protein), and "Protein BLAST" (protein to protein). Each option is represented by a blue button with a corresponding icon (DNA helix for Nucleotide BLAST, a protein ribbon for Protein BLAST, and a central button for blastx/tblastn).

## 1.2. Select “Nucleotide BLAST”:

The screenshot shows the NCBI BLAST suite interface. At the top, there are logos for NIH (U.S. National Library of Medicine) and NCBI (National Center for Biotechnology Information). Below the logos, the text "BLAST® >> blastn suite" is displayed. The main heading is "Standard Nucleotide BLAST". There are tabs for "blastn", "blastp", "blastx", "tblastn", and "tblastx", with "blastn" being the active tab. The interface is divided into two main sections: "Enter Query Sequence" and "Choose Search Set". In the "Enter Query Sequence" section, there is a large text area for "Enter accession number(s), gi(s), or FASTA sequence(s)", a "Clear" button, and a "Query subrange" section with "From" and "To" input fields. Below this, there is a section for "Or, upload file" with a "Choose File" button and "No file chosen" text. There is also a "Job Title" input field with a placeholder "Enter a descriptive title for your BLAST search". A checkbox labeled "Align two or more sequences" is present. In the "Choose Search Set" section, there is a "Database" section with radio buttons for "Standard databases (nr etc.)", "rRNA/ITS databases", and "Genomic + transcript databases". The "Standard databases (nr etc.)" radio button is selected, and a dropdown menu shows "Nucleotide collection (nr/nt)". There is also an "Organism" section with a text input field "Enter organism name or id—completions will be suggested" and an "exclude" checkbox with a "+" button.

## 1.3. Paste “Sequence A” in the form;

In the menu “Database” select “Genomic + transcript databases”;

In the drop-down menu under “Database” select “Human genomic plus transcript (G+T)”:

This screenshot shows the same NCBI BLAST suite interface as the previous one, but with more data entered. In the "Enter Query Sequence" section, the "Enter accession number(s), gi(s), or FASTA sequence(s)" text area now contains a multi-line FASTA sequence: "AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGAAGAATTTCTGTTCTCAGTTTCTGGATTATG CCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTCCTATGATGAATATAGATACAGAAGCGTCAAGCATGCCA ACTAGAAGAGGTAAGAAGAACTATGTGAAAACCTTTTGATTATGCATATGAAC". The "Query subrange" section still has empty "From" and "To" fields. In the "Choose Search Set" section, the "Database" radio buttons are now: "Standard databases (nr etc.)", "rRNA/ITS databases", and "Genomic + transcript databases". The "Genomic + transcript databases" radio button is now selected. The dropdown menu for "Database" now shows "Human genomic plus transcript (Human G+T)". Below this, there is an "Exclude" section with checkboxes for "Models (XM/XP)" and "Uncultured/environmental sample sequences". There is also a "Limit to" section with a checkbox for "Sequences from type material". Below that, there is an "Entrez Query" section with a text input field "Enter an Entrez query to limit search" and a "YouTube" button. At the bottom, there is a "Program Selection" section with a radio button for "Highly similar sequences (megablast)".

1.4. In the section “Program selection”, select “Highly similar sequences (megablast)”:

Choose Search Set

Database

☐ Standard databases (nr etc.):
 ☐ rRNA/ITS databases
 ☒ Genomic + transcript databases

☒ Human genomic plus transcript (Human G+T)

Exclude  
Optional

☐ Models (XM/XP)
 ☐ Uncultured/environmental sample sequences

Limit to  
Optional

☐ Sequences from type material

Entrez Query  
Optional

[YouTube](#) [Create custom database](#)

Enter an Entrez query to limit search

Program Selection

Optimize for

☒ Highly similar sequences (megablast)
 ☐ More dissimilar sequences (discontiguous megablast)
 ☐ Somewhat similar sequences (blastn)

Choose a BLAST algorithm

BLAST

Search **database Human G+T** using **Megablast (Optimize for highly similar sequences)**

☐ Show results in a new window

1.5. Now all parameters are selected, and we can press the “BLAST” button to start analysis:

NIH

U.S. National Library of Medicine

NCBI

National Center for Biotechnology Information

**BLAST**
>> blastn suite
>> RID-1PDNXS4H014

Format Request Status

[\[Formatting options\]](#)

Job Title: Nucleotide Sequence

Request ID	1PDNXS4H014
Status	Searching
Submitted at	Sun Jan 12 10:09:49 2020
Current time	Sun Jan 12 10:09:51 2020
Time since submission	00:00:02

This page will be automatically updated in 2 seconds

1.6. When the program finishes the analysis you will see the header like this:

**BLAST® » blastn suite » results for RID-1PDNXS4H014**

[< Edit Search](#)
[Save Search](#)
[Search Summary ▾](#)

<b>Job Title</b>	<b>Nucleotide Sequence</b>
<b>RID</b>	<a href="#">1PDNXS4H014</a> <span>Search expires on 01-13 22:09 pm</span> <a href="#">Download All ▾</a>
<b>Program</b>	BLASTN <a href="#">?</a> <a href="#">Citation ▾</a>
<b>Database</b>	Human G+T (2 databases) <a href="#">See details ▾</a>
<b>Query ID</b>	lcl Query_42429
<b>Description</b>	None
<b>Molecule type</b>	dna
<b>Query Length</b>	203
<b>Other reports</b>	<a href="#">Distance tree of results</a> <a href="#">MSA viewer</a> <a href="#">?</a>

Descriptions

Graphic Summary

Alignments

Taxonomy

Scroll down to the most important part of this page:

Program BLASTN [?](#) [Citation ▾](#)  
Database Human G+T (2 databases) [See details ▾](#)  
Query ID lcl|Query\_42429  
Description None  
Molecule type dna  
Query Length 203  
Other reports [Distance tree of results](#) [MSA viewer](#) [?](#)

Organism only top 20 will appear [Exclude](#)  
  
[+ Add organism](#)  

Percent Identity  to   
E value  to   
Query Coverage  to   
[Filter](#) [Reset](#)

Descriptions

Graphic Summary

Alignments

Taxonomy

Sequences producing significant alignments
Download ▾ Manage Columns ▾ Show 100 ▾ [?](#)

☒ select all 2 sequences selected
[GenBank](#) [Graphics](#) [Distance tree of results](#)

	Description	Max Score	Total Score	Query Cover	E value	Per. Ident	Accession
Transcripts							
<input checked="" type="checkbox"/>	<a href="#">Homo sapiens cystic fibrosis transmembrane conductance regulator (CFTR). mRNA</a>	283	283	80%	3e-74	97.60%	<a href="#">NM_000492.3</a>

In this case, the DNA sequence A that we submitted has mapped to the human gene *CFTR* (cystic fibrosis transmembrane conductance regulator). The E-value is 3e-74 – the meaning of this parameter can be roughly understood as the probability to obtain the same result by chance (that is, if we would randomly construct a DNA sequence of 208 nucleotides, the probability that it would map to the same gene with the same similarity would be 3e-74). Thus, the chance that this result would be obtained by a random coincidence is very small, or in other words, the statistical significance of this result is very high.

2. If sequence A mapped to a known human gene, check in the BLAST output whether mutations are present in this gene.

Let's click on the line "Homo sapiens cystic fibrosis transmembrane conductance (CFTR), mRNA":

Descriptions

Graphic Summary

Alignments

Taxonomy

Alignment view

Pairwise

CDS feature

2 sequences selected

Download

GenBank

Graphics

Homo sapiens cystic fibrosis transmembrane conductance regulator (CFTR), mRNA

Sequence ID: [NM\\_000492.3](#) Length: 6132 Number of Matches: 1

Range 1: 1551 to 1717 [GenBank](#) [Graphics](#) [Next Match](#) [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
283 bits(153)	3e-74	163/167(98%)	4/167(2%)	Plus/Plus
Query 1	AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGGAGAATTTATTCTGTTCTCA	60		
Sbjct 1551	AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGGAGAATTTATTCTGTTCTCA	1610		
Query 61	GTTTTCTGGATTATGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTTCCTATGA	120		
Sbjct 1611	GTTTTCTGGATTATGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTTCCTATGA	1670		
Query 121	TGAATATAGATACAGAAGCGTC----AAGCATGCCAACTAGAAGAGG	163		
Sbjct 1671	TGAATATAGATACAGAAGCGTCATCAAAGCATGCCAACTAGAAGAGG	1717		

This graph shows the alignment of "Sequence A" to the *CFTR* gene in the human genome. As you can see from this graph, only four nucleotides do not match ("Sequence A" has a deletion of these four nucleotides which appear in the reference human genome but do not appear in "Sequence A"). The overall identity between these two sequences is 98%, which is a very good match (highly unlikely to come up with such sequence randomly by chance). The four nucleotides which are missing represent a mutation (deletion).

3. Check how this DNA sequence translates into amino-acid sequence using ExPASy:

3.1. Go to the ExPASy web site: <https://web.expasy.org/translate/>

3.2. Paste the DNA sequence A in the form:

**Translate** is a tool which allows the translation of a nucleotide (DNA/RNA) sequence to a protein sequence.

**DNA or RNA sequence**

```
AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGGAAGAATTTTCATTCTGTTCTCAGTTTTCTGGATTA
TGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTCCTATGATGAATATAGATACAGAAAGCGTCAAGCAT
GCCAACTAGAAGAGGTAAGAAACTATGTGAAAACTTTTTGATTATGCATATGAAC
```

**Output format**

- ☐ Verbose: Met, Stop, spaces between residues
- ☒ Compact: M, -, no spaces
- ☐ Includes nucleotide sequence
- ☐ Includes nucleotide sequence, no spaces

**DNA strands**

☒ forward ☒ reverse

**Genetic codes** - [See NCBI's genetic codes](#)

Standard

reset TRANSLATE!

3.3. Get resulting amino-acid sequences for different reading frames:

**Results of translation**

- Open reading frames are highlighted in red
- Select your initiator on one of the following frames to retrieve your amino acid sequence

Download all the translated frames

**5'3' Frame 1**

RTGAFRG-N-AQWKNFIFSFLDYAWHH-RKYHLWCFL--I-IQKRQACQLEEVNRYVKTFLCI-

**5'3' Frame 2**

ELPSEGGIKHSGRISFCQFSWIMPGTIKENIIFGVSYDEYRYSVKHAN-KR-ETM-KLFDYAYE

**5'3' Frame 3**

NWSLQRVKLSTVEEFHVSLSFPLCLAPLKKISSLVFMMNIDTEASSMPTRRGKKLCENFLIMHM

**3'5' Frame 1**

VHMHQKVFT-FLTSSSWHA-RFCYIHHRRKHQR-YFL-WCQA-SRKTENRMKFFHCA-FYPLKAPV

**3'5' Frame 2**

FICIIKKFHSFLPLLVGMLDASVSIFIIGNTKDDIFFNGARHNPGLRTE-NSSTVLNFTL-RLQF

**3'5' Frame 3**

SYA-SKSFHIVSYLF-LACLTLLYLYSS-ETPRMIFSLMVPGLIQEN-EQNEILPLCLILPSEGSSS

3.4. Now let's compare with the wild-type sequence in the reference DNA genome. You can get this sequence from the BLAST alignment output as shown in the figure below:

#### Homo sapiens cystic fibrosis transmembrane conductance regulator (CFTR), mRNA

Sequence ID: [NM\\_000492.3](#) Length: 6132 Number of Matches: 1

Range 1: 1551 to 1717 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Pre

Score	Expect	Identities	Gaps	Strand
283 bits(153)	3e-74	163/167(98%)	4/167(2%)	Plus/Plus
Query 1	AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGAAGAATTTCACTGTTCTCA	60		
Sbjct 1551	AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGAAGAATTTCACTGTTCTCA	1610		
Query 61	GTTTTCTGGATTATGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTCCTATGA	120		
Sbjct 1611	GTTTTCTGGATTATGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTCCTATGA	1670		
Query 121	TGAATATAGATACAGAAGCGTC----AAGCATGCCAACTAGAAGAGG	163		
Sbjct 1671	TGAATATAGATACAGAAGCGTCATCAAGCATGCCAACTAGAAGAGG	1717		

3.5. Copy the wild-type DNA sequence (it is shown in the red rectangle in the figure below):

#### Homo sapiens cystic fibrosis transmembrane conductance regulator (CFTR), mRNA

Sequence ID: [NM\\_000492.3](#) Length: 6132 Number of Matches: 1

Range 1: 1551 to 1717 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Pre

Score	Expect	Identities	Gaps	Strand
283 bits(153)	3e-74	163/167(98%)	4/167(2%)	Plus/Plus
Query 1	AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGAAGAATTTCACTGTTCTCA	60		
Sbjct 1551	AGAACTGGAGCCTTCAGAGGGTAAAATTAAGCACAGTGAAGAATTTCACTGTTCTCA	1610		
Query 61	GTTTTCTGGATTATGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTCCTATGA	120		
Sbjct 1611	GTTTTCTGGATTATGCCTGGCACCATTAAAGAAAATATCATCTTTGGTGTTCCTATGA	1670		
Query 121	TGAATATAGATACAGAAGCGTC----AAGCATGCCAACTAGAAGAGG	163		
Sbjct 1671	TGAATATAGATACAGAAGCGTCATCAAGCATGCCAACTAGAAGAGG	1717		

3.6. Paste the wild-type DNA sequence to ExPASy (<https://web.expasy.org/translate/>) and translate it to the amino-acid sequence for all possible reading frames:

**Results of translation**

- Open reading frames are highlighted in red
- Select your initiator on one of the following frames to retrieve your amino acid sequence

[Download all the translated frames](#)

**5'3' Frame 1**

RTGAFRG-N-AQWKNFILSVFLDYAWHH-RKYHLWCFL--I-IQKRHQSMPTRB

**5'3' Frame 2**

ELPSEGKIKHSGRISFCSQFSWIMPGTIKENIIFGVSYDEYRYSVIKACQLEE

**5'3' Frame 3**

NWSLQVRVKLSTVEEFHVSLSFPGLCIAPLKKISSLVFPMNIDTEASSKHAN-KR

**3'5' Frame 1**

PLLVGML--RFCIYIHRKHQR-YFL-WQA-SRKTENRMKFFHCA-FYPLKAPV

**3'5' Frame 2**

LF-LACFDASVSIFIIGNTKDDIFFNGARHNPGKLRT-NSSTVLNFTL-RLQF

**3'5' Frame 3**

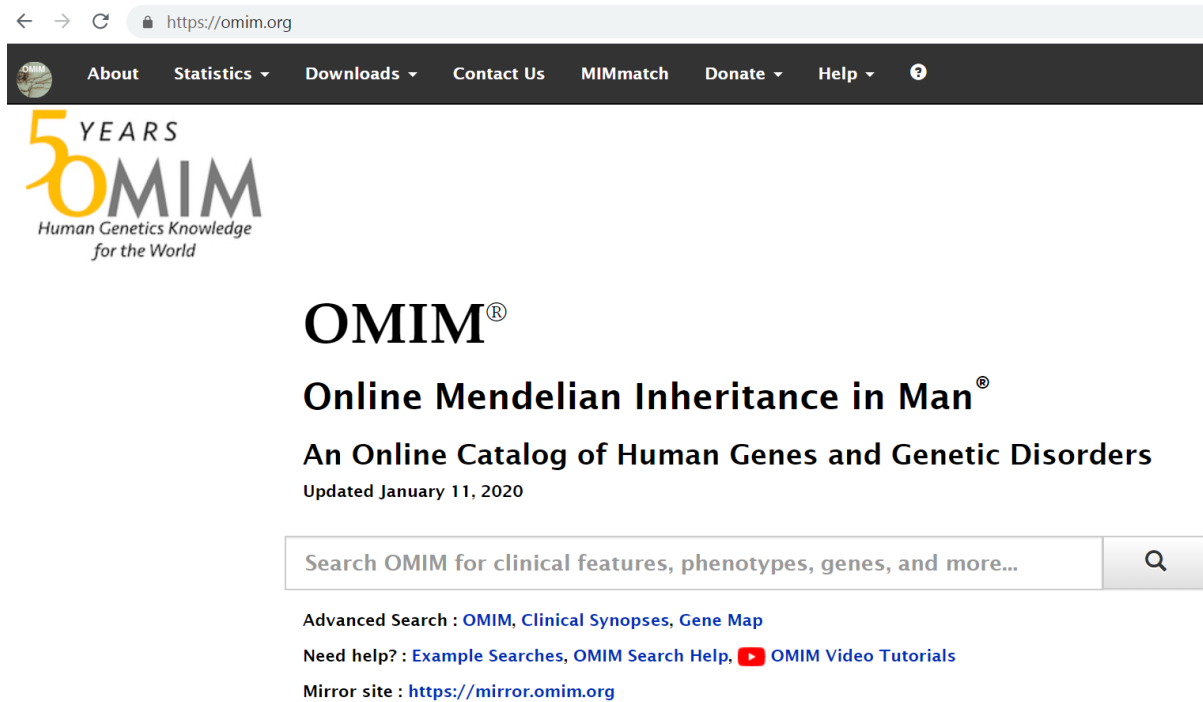
SSSWHALMTLLLYSS-ETPKMIFSLMVGIIQEN-EQNEILPLCLILPSEGSSS



### 3.7. Discuss: what amino-acid changes can be caused by this mutation?

4. Check in the OMIM database, which Mendelian disease is associated with this mutation.

4.1. Let's go to the OMIM database: <https://omim.org>



← → ↺ <https://omim.org>

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Human Genetics Knowledge for the World

**OMIM®**  
**Online Mendelian Inheritance in Man®**  
An Online Catalog of Human Genes and Genetic Disorders  
Updated January 11, 2020

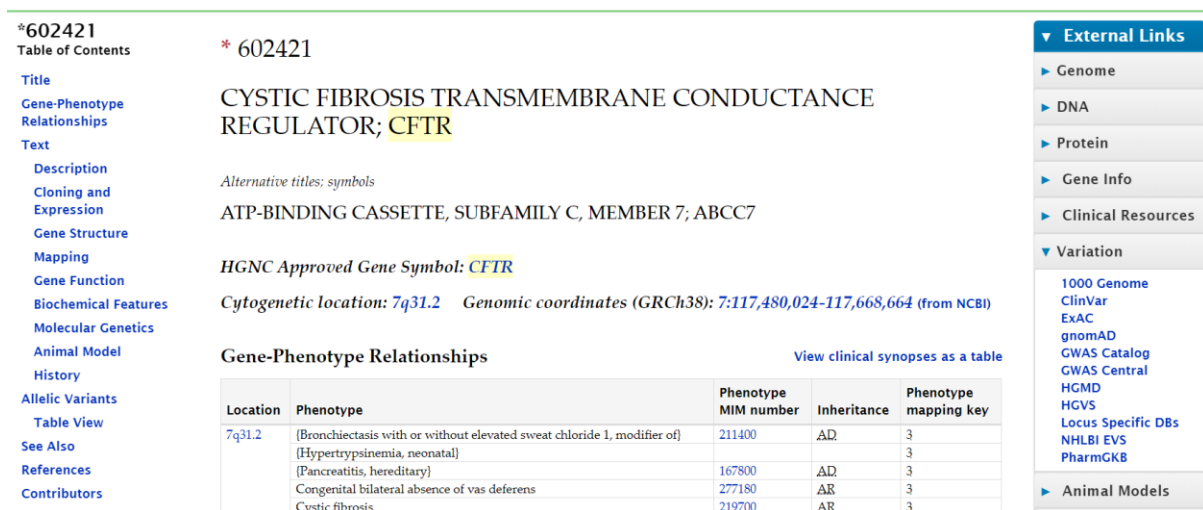
Search OMIM for clinical features, phenotypes, genes, and more...

Advanced Search : [OMIM](#), [Clinical Synopses](#), [Gene Map](#)

Need help? : [Example Searches](#), [OMIM Search Help](#), [OMIM Video Tutorials](#)

Mirror site : <https://mirror.omim.org>

4.2. Let's look for the *CFTR* gene in the OMIM database:



\*602421  
Table of Contents

Title  
Gene-Phenotype Relationships  
Text  
Description  
Cloning and Expression  
Gene Structure  
Mapping  
Gene Function  
Biochemical Features  
Molecular Genetics  
Animal Model  
History  
Allelic Variants  
Table View  
See Also  
References  
Contributors

\* 602421  
CYSTIC FIBROSIS TRANSMEMBRANE CONDUCTANCE REGULATOR; **CFTR**

Alternative titles; symbols  
ATP-BINDING CASSETTE, SUBFAMILY C, MEMBER 7; ABCC7

HGNC Approved Gene Symbol: **CFTR**

Cytogenetic location: **7q31.2** Genomic coordinates (GRCh38): **7:117,480,024-117,668,664** (from NCBI)

Gene-Phenotype Relationships [View clinical synopses as a table](#)

Location	Phenotype	Phenotype MIM number	Inheritance	Phenotype mapping key
7q31.2	[Bronchiectasis with or without elevated sweat chloride 1, modifier of]	211400	AD	3
	[Hypertrypsinemia, neonatal]			3
	[Pancreatitis, hereditary]	167800	AD	3
	Congenital bilateral absence of vas deferens	277180	AR	3
	Cystic fibrosis	219700	AR	3

**External Links**

- Genome
- DNA
- Protein
- Gene Info
- Clinical Resources
- Variation
  - 1000 Genome
  - ClinVar
  - ExAC
  - gnomAD
  - GWAS Catalog
  - GWAS Central
  - HGMD
  - HGVS
  - Locus Specific DBs
  - NHLBI EVS
  - PharmGKB
- Animal Models

4.3. Work independently with this page of the OMIM database to read about possible phenotypes associated with CFTR gene and associated medical information.

**Discuss: is cystic fibrosis a recessive or dominant disease? If this patient has a piece of DNA with mutation in CFTR, does it mean she/he has cystic fibrosis? What are its molecular mechanisms?**

## 5\*. Try to map sequence B to the human genome using BLAST

Repeat steps 1.1-1.6 to map “Sequence B” to the human genome.

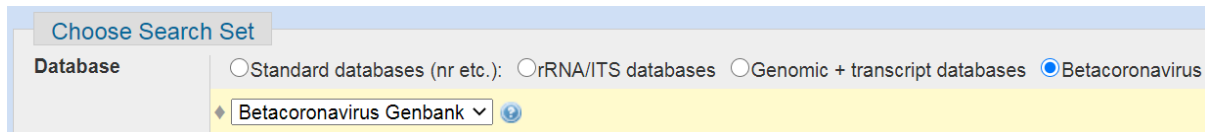
Here is “sequence B”:

```
ATTTGATCCTGGCTCAGGACGAACGCTGGCGGCGTGCCTAATACATGCAAGTAGAACGCTGAAGGAGGAG
CTTGCTTCTCTGGATGAGTTGCGAACGGGTGAGTAACGCGTAGGTAACCTGCCTGGTAGCGGGGGATAAC
TATTGGAAACGATAGCTAATACCGCATAAGAGTGGATGTTGCATGACATTTGCTTAAAAGGTGCACTTGC
ATCACTACCAGATGGACCTGCGTTGTATTAGCTAGTTGGTGGGGTAACGGCTCACCAAGGCGACGATACA
TAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGKCCCAGACTCCTACGGGAGGCAGCA
```

**Discuss: Did you manage to map “Sequence B” to the human genome? Why?**

## 6. Try to map sequence B to the coronavirus genome using BLAST

Repeat steps 1.1-1.6, but select “Betacoronavirus” as the database:



**Discuss: Did you manage to map “Sequence B” to the coronavirus genome? Why?**

**BTW, it’s possible to say that it’s not RNA from virus even without this analysis! Why?**

## 7. Map “Sequence B” to bacterial and fungal genomes

7.1. Open BLAST (<https://blast.ncbi.nlm.nih.gov>) and in the section “Database” select “rRNA/ITS databases”. In the drop-down menu select “16S ribosomal RNA sequences (bacteria and fungi)”<sup>ii</sup>:

blastn blastp blastx tblastn tblastx

BLASTN programs search nucleotide databases using a nucleotide query sequence

### Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [?](#) [Clear](#) [Query subrange](#) [?](#)

From

To

ATTGATCCTGGCTCAGGACGAACGCTGGCGGCTGCCTAATACATGCAAGTAGAACGCTGAAGGAGGAG  
CTTGCTTCTCTGGATGAGTTGCGAACGGGTGAGTAACGCGTAGGTAACCTGCCTGGTAGCGGGGATAAC  
TATTGGAAACGATAGCTAATACCGCATAAGAGTGGATGTTGCATGACATTTGCTTAAAGGTGCACCTTGC  
ATCACTACCAGATGGACCTGCGTTGTATTAGCTAGTTGGTGGGTAAACGGCTCACCAGGCGACGATACA  
TAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGKCCAGACTCCTACGGGAGGCAGCA

Or, upload file  No file chosen [?](#)

Job Title

Enter a descriptive title for your BLAST search [?](#)

☐ Align two or more sequences [?](#)

### Choose Search Set

Database ☐ Standard databases (nr etc.): ☒ rRNA/ITS databases ☐ Genomic + transcript databases

16S ribosomal RNA sequences (Bacteria and Archaea) [?](#)

Organism [Optional](#)  ☐ exclude [+](#)

Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown [?](#)

Exclude [Optional](#) ☐ Models (XM/XP) ☐ Uncultured/environmental sample sequences

Limit to [Optional](#) ☐ Sequences from type material

Entrez Query [Optional](#)  [YouTube](#) [Create custom database](#)

### Program Selection

Optimize for ☒ Highly similar sequences (megablast)

Click button “BLAST”:

[BLAST](#) Search database 16S ribosomal RNA sequences (Bacteria and Archaea) using Megablast (Optimize for highly similar sequences)

☐ Show results in a new window

[+ Algorithm parameters](#) [Note: Parameter values that differ from the default are highlighted in yellow and marked with ♦ sign](#)

7.2. After the program finishes calculations, you will get the following results:

Program [BLASTN](#) [Citation](#) [?](#)

Database rRNA\_typestrains/prokaryotic\_16S\_ribosomal\_RNA [See details](#) [?](#)

Query ID lcl|Query\_7965

Description None

Molecule type dna

Query Length 350

Other reports [Distance tree of results](#) [MSA viewer](#) [?](#)

Organism [only top 20 will appear](#) ☐ exclude

[+ Add organism](#)

Percent Identity  to

E value  to

Query Coverage  to

[Filter](#) [Reset](#)

**Descriptions** [Graphic Summary](#) [Alignments](#) [Taxonomy](#)

**Sequences producing significant alignments** [Download](#) [Manage Columns](#) [Show](#)  [?](#)

☒ select all 100 sequences selected [GenBank](#) [Graphics](#) [Distance tree of results](#)

	Description	Max Score	Total Score	Query Cover	E value	Per. Ident	Accession
<input checked="" type="checkbox"/>	<a href="#">Streptococcus pneumoniae strain ATCC 33400 16S ribosomal RNA, partial sequence</a>	643	643	100%	0.0	100.00%	NR_028865.1
<input checked="" type="checkbox"/>	<a href="#">Streptococcus mitis strain NS51 16S ribosomal RNA, partial sequence</a>	625	625	99%	6e-179	98.85%	NR_028864.1
<input checked="" type="checkbox"/>	<a href="#">Streptococcus pneumoniae strain ATCC 33400 16S ribosomal RNA, partial sequence</a>	612	612	95%	4e-175	100.00%	NR_117496.1

7.3. Click on the top match to see how “Sequence B” aligned with it:

### Streptococcus pneumoniae strain ATCC 33400 16S ribosomal RNA, partial sequence

Sequence ID: [NR\\_028665.1](#) Length: 1515 Number of Matches: 1

Range 1: 1 to 350 [GenBank](#) [Graphics](#)

▼ [Next Match](#) ▲ [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
643 bits(348)	0.0	350/350(100%)	0/350(0%)	Plus/Plus
Query 1	ATTTGATCCTGGCTCAGGACGAACGCTGGCGGCGTGCCTAATACATGCAAGTAGAACGCT	60		
Sbjct 1	ATTTGATCCTGGCTCAGGACGAACGCTGGCGGCGTGCCTAATACATGCAAGTAGAACGCT	60		
Query 61	GAAGGAGGAGCTTGCTTCTCTGGATGAGTTGCGAACGGGTGAGTAACGCGTAGGTAACCT	120		
Sbjct 61	GAAGGAGGAGCTTGCTTCTCTGGATGAGTTGCGAACGGGTGAGTAACGCGTAGGTAACCT	120		
Query 121	GCCTGGTAGCGGGGGATAACTATTGGAAACGATAGCTAATACCGCATAAGAGTGGATGTT	180		
Sbjct 121	GCCTGGTAGCGGGGGATAACTATTGGAAACGATAGCTAATACCGCATAAGAGTGGATGTT	180		
Query 181	GCATGACATTTGCTTAAAAGGTGCACTTGCACTACTACCAGATGGACCTGCGTTGTATTA	240		
Sbjct 181	GCATGACATTTGCTTAAAAGGTGCACTTGCACTACTACCAGATGGACCTGCGTTGTATTA	240		
Query 241	GCTAGTTGGTGGGGTAACGGCTCACCAAGGCGACGATACATAGCCGACCTGAGAGGGTGA	300		
Sbjct 241	GCTAGTTGGTGGGGTAACGGCTCACCAAGGCGACGATACATAGCCGACCTGAGAGGGTGA	300		
Query 301	TCGGCCACACTGGGACTGAGACACGKCCCAGACTCCTACGGGAGGCAGCA	350		
Sbjct 301	TCGGCCACACTGGGACTGAGACACGKCCCAGACTCCTACGGGAGGCAGCA	350		

8. As you can see from this output, the piece of DNA extracted from the blood of the patient belongs to *Streptococcus pneumoniae* strain ATCC 33400

**8.1. Discuss: How can it be that the cell-free DNA fraction in the blood plasma contains this piece of DNA that maps to a pathogenic bacterium?**

**8.2. Discuss: What would you advise to medical doctors?**

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<sup>i</sup> In a real situation sequencing of cell-free DNA would return millions of short pieces of DNA and we would need to do more advanced analysis, but for the purpose of this practical for simplicity only two DNA sequences were reported. For example, this could be the result of targeted amplification of DNA sequences of interest.

<sup>ii</sup> rRNA genes are extremely conserved across many bacterial and fungal species, therefore rRNA is frequently used in cross-species mapping. While rRNA is very conserved, there are differences between different species, so if a given sequence of rRNA is compared to rRNA from each known species of bacteria and fungi it is possible to identify the best match. Thus, we can uniquely identify the bacteria or fungi to which a given piece of DNA belongs.